

IRIDOID AND PHENOLIC GLUCOSIDE FROM *VITEX ROTUNDIFOLIA*

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Key Word Index—*Vitex rotundifolia*; Verbenaceae; iridoid glucoside; agnuside; eurostoside; phenylbutanone glucoside.

Abstract—The phenylbutanone glucoside, as well as the known compounds agnuside and eurostoside, were isolated from the leaves of *Vitex rotundifolia*, although eurostoside was obtained as a *trans*- and *cis*-mixture. The structures were established on the basis of spectroscopic data.

INTRODUCTION

Vitex rotundifolia L.f. is widely distributed on the sea coast in Asia. The presence of the iridoid glucoside, agnuside [1], and the diterpene, rotundifuran [2] have been recorded. This report deals with a new constituent of *V. rotundifolia*.

RESULTS AND DISCUSSION

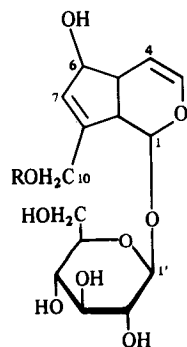
From the methanol extract of the leaves of *V. rotundifolia*, two iridoid glucosides **1** and **2** and a phenol glucoside **3** were isolated.

The ^1H and ^{13}C NMR spectra of compound **1** were identical with those of agnuside [3, 4], which was isolated as a major component from this plant [1].

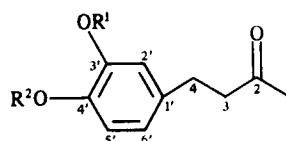
Compound **2** analysed for $\text{C}_{24}\text{H}_{28}\text{O}_{11}$ [FABMS, m/z 515 $[\text{M} + \text{H}]^+$]. The ^1H NMR spectrum of compound **2** is very similar to that of agnuside except for the signals of the aromatic and olefinic protons. Although these low field signals are too complex to clarify, two sets of *para*-substituted cinnamoyl moieties were elucidated by the proton–proton decoupling experimental, i.e. the *trans*-olefin system ($J = 16.1$ Hz) at δ 6.36 and 7.65 (each doublet), and the *cis*-olefin system ($J = 12.6$ Hz) at 5.81 and 6.90 (each doublet) were found, along with the *para*-substituted benzene ring protons (7.46, 6.80, each *d*, $J = 8.7$ Hz, 1H; 7.63, 6.75, each *d*, $J = 8.9$ Hz, 1H). As the compound **2** gave a hexa-acetate (**2a**) (EIMS, m/z 744 $[\text{M}]^+$), these findings indicated **2** has a *p*-hydroxycinnamoyl moiety instead of the *p*-hydroxybenzoyl found in agnuside (**1**). The position of the cinnamoyl moiety was established as C-10, as the signals of the glucose moiety in the ^{13}C NMR spectrum and the signals of H-6 (δ 4.45) and H-10 (δ 4.70) in the ^1H NMR spectrum of **2** were similar to those of agnuside (**1**) [3, 4]. It was concluded that **2** is a 1:1 mixture of 10-*O-trans*- and *cis-p*-hydroxycinnamoyl aucubin. Compound **2** was separated into *cis*- and *trans*-compounds by HPLC, but it was still a mixture after

leaving it at room temperature. The *trans*-compound **2** is known from *Euphrasia rostkoviana* [5], *Penstemon whippleanus* [6], and *Vitex agnuscastus* [7], and named eurostoside.

Compound **3** analysed for $\text{C}_{16}\text{H}_{22}\text{O}_8$ (FABMS, m/z 365 $[\text{M} + \text{Na}]^+$). The keto group was suggested by an IR absorption at 1700 cm^{-1} , and a methyl ketone signal at δ 2.11 in the ^1H NMR spectrum. Moreover, the ^1H NMR spectrum of **3** showed four equivalent protons at δ 2.74 (*br s*, 4H) and the trisubstituted benzene ring protons at 6.60 (*dd*, $J = 10.6$ and 2.2 Hz, 1H), 6.69 (*d*, $J = 2.2$ Hz, 1H) and 7.07 (*d*, $J = 10.6$ Hz, 1H). The ^{13}C NMR of **3** indicated the



- 1** R = *p*-hydroxybenzoyl
2 R = *trans* and *cis* *p*-hydroxycinnamoyl



- 3** R¹ = H, R² = Glu
3a R¹ = Ac, R² = Glu(OAc)₄
4 R¹ = Glu, R² = H

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Table 1. ^{13}C NMR data of myzodendrone* and compound 3 (22.5 MHz)

Position <i>n</i> -chain	Myzodendrone	3
C-1	30.2 <i>q</i>	30.0 <i>q</i>
C-2	165.6 <i>s</i> †	211.4 <i>s</i>
C-3	45.9 <i>t</i>	45.6 <i>t</i>
C-4	30.2 <i>t</i>	30.0 <i>t</i>
Benzene ring		
C-1'	124.5 <i>s</i>	138.3 <i>s</i>
C-2'	117.0 <i>d</i>	117.0 <i>d</i>
C-3'	134.2 <i>s</i>	144.8 <i>s</i>
C-4'	146.5 <i>s</i>	148.2 <i>s</i>
C-5'	118.9 <i>d</i>	119.3 <i>d</i>
C-6'	119.0 <i>d</i>	120.7 <i>d</i>
Glucose		
C-1''	104.3 <i>d</i>	104.5 <i>d</i>
C-2''	74.9 <i>d</i>	74.7 <i>d</i>
C-3''	77.6 <i>d</i>	77.5 <i>d</i>
C-4''	71.4 <i>d</i>	71.1 <i>d</i>
C-5''	78.3 <i>d</i>	78.0 <i>d</i>
C-6''	62.5 <i>t</i>	62.4 <i>t</i>

All values are in (ppm) relative to TMS (CD_3OD).

*Data obtained from ref. [8].

†This assignment is thought to be incorrect.

presence of a benzene ring, two methylene carbons and a sugar group which was confirmed as glucose by chromatographic comparison of the hydrolysis product of 3 with an authentic sample. Compound 3 gave a penta-acetate (3a) upon usual acetylation. All these findings indicated 3 is 3',4'-dihydroxyphenylbutanone glucoside in comparison with myzodendrone (4) [4-(3',4'-dihydroxyphenyl)-butan-2-one-3'-*O*-glucoside] which was isolated from *Myzodendron punctulatum* [8].

The position of glucose was established as 4' by the NOE enhancement (10%) of the signal at $\delta 7.07$ (H-5') of the aromatic proton upon irradiation at $\delta 4.68$ (anomeric proton, H-1'') frequency. The β -configuration of C-1'' (anomeric carbon) was deduced by the signals of glucose in the ^{13}C NMR spectrum of 3 and the *J* value (*J* = 7.5 Hz) at $\delta 4.68$ of the anomeric proton signal in the ^1H NMR spectrum of 3. Thus, it was concluded that compound 3 is 4-(3',4'-dihydroxyphenyl)-butan-2-one-4'-*O*- β -D-glucoside.

EXPERIMENTAL

^1H and ^{13}C NMR spectra were recorded in CD_3OD solns unless otherwise specified and 90.0/22.5 MHz ($^1\text{H}/^{13}\text{C}$) using TMS as int. standard.

Extraction and isolation. The dried leaves of *V. rotundifolia* were collected on the sea coast in Nagasaki, Japan, and extracted

with MeOH. The MeOH extracts (362 g) were dissolved in H_2O , and partitioned between *n*-hexane and H_2O , then *n*-BuOH and H_2O , successively. The *n*-BuOH soluble portion was extracted with EtOAc–MeOH (7:3) to give the soluble part (42.3 g). This was chromatographed on silica gel using the solvent system of CHCl_3 –MeOH– H_2O (8:2:0.1), then purified by reversed phase chromatography [solvent system MeOH– H_2O (6:4)] to give the compounds 1 (23 g), 2 (345 mg) and 3 (290 mg), respectively. The ^1H and ^{13}C NMR spectra of 1 were identical with those of agnuside.

Compound 2. Amorphous powder, $[\alpha]_D^{16} -49.6^\circ$ (EtOH; *c* 0.9); FABMS *m/z* 515 $[\text{M} + \text{Na}]^+$; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3550, 1700 and 1610; ^1H NMR: δ 5.00 (*d*, *J* = 7.4 Hz, 1H, H-1), 6.39 (*dd*, *J* = 2.0, 6.3 Hz, 1H, H-3), 5.11 (*dd*, *J* = 3.8, 6.3 Hz, 1H, H-4), 2.70 (*m*, 1H, H-5), 4.45 (*m*, 1H, H-6), 5.80 (*br s*, 1H, H-7), 2.96 (*br t*, *J* = 7.4 Hz, 1H, H-9), 4.70 (*d*, *J* = 7.1 Hz, 2H, H-10), 4.69 (*d*, *J* = 7.1 Hz, 1H, H-1'), *p*-hydroxy cinnamoyl moiety; H-*trans* 6.36 (*d*, *J* = 16.1 Hz, 1H), 7.65 (*d*, *J* = 16.1 Hz, 1H), 7.46 (*d*, *J* = 8.7 Hz, 2H), 6.80 (*d*, *J* = 8.7 Hz, 2H); H-*cis* 5.81 (*d*, *J* = 12.6 Hz, 1H), 6.90 (*d*, *J* = 12.6 Hz, 1H), 7.63 (*d*, *J* = 8.9 Hz, 2H), 6.75 (*d*, *J* = 8.9 Hz, 2H); ^{13}C NMR: glucose moiety δ 100.3 (*d*, C-1'), 74.9 (*d*, C-2'), 78.2 (*d*, C-3'), 71.5 (*d*, C-4'), 78.0 (*d*, C-5') and 62.8 (*t*, C-6').

Compound 3. Amorphous powder. (Found: C, 52.41; H, 6.54. $\text{C}_{16}\text{H}_{22}\text{O}_8 \cdot 3/2\text{H}_2\text{O}$ requires: C, 52.03; H, 6.83%). $[\alpha]_D^{16} -39.6^\circ$ (MeOH; *c* 0.4); FABMS *m/z* 365 $[\text{M} + \text{Na}]^+$; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3500–3100, 1702, 1600 and 1510; ^1H NMR: δ 2.11 (*s*, 3H, H-1), 2.74 (*br s*, 4H, H-3 and 4), 3.35–3.90 (*m*, 6H, sugar protons), 4.68 (*d*, *J* = 7.5 Hz, 1H, anomeric proton H-1''), 6.60 (*dd*, *J* = 10.6 and 2.2 Hz, 1H, H-6'), 6.99 (*d*, *J* = 2.2 Hz, 1H, H-2'), 7.07 (*d*, *J* = 10.6 Hz, 1H, H-5').

Acetate of compound 3. Prepared with Ac_2O –pyridine; 3a colourless oil, $[\alpha]_D^{16} -11.2^\circ$ (CHCl_3 ; *c* 0.3); EIMS *m/z* 552 $[\text{M}]^+$; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1745, 1713, 1600 and 1509; ^1H NMR (CDCl_3): δ 2.03, 2.04, 2.07 and 2.08 (each *s*, 3H, acetyl signals), 2.12 (*s*, 3H, H-1), 2.25 (*s*, 3H, 3'-OAc), 2.79 (*m*, 4H, H-3 and 4), 3.96 (*d*, *J* = 7.1 Hz, 1H, H-1''), 4.22 (*m*, 2H, H-6''), 5.00–5.34 (*m*, 4H, sugar protons), 6.87–6.96 (*m*, 3H, H-2', 5' and 6').

Acid hydrolysis of compound 3. Compound 3 (30 mg) was refluxed with 0.1 N HCl (15 ml) for 30 min, the mixture was neutralized with $\text{Ba}(\text{OH})_2$ and filtered. The filtrate was evapd and the residue was examined by Avicel TLC in comparison with the authentic sample of glucose.

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